

Epitope-focused vaccine design

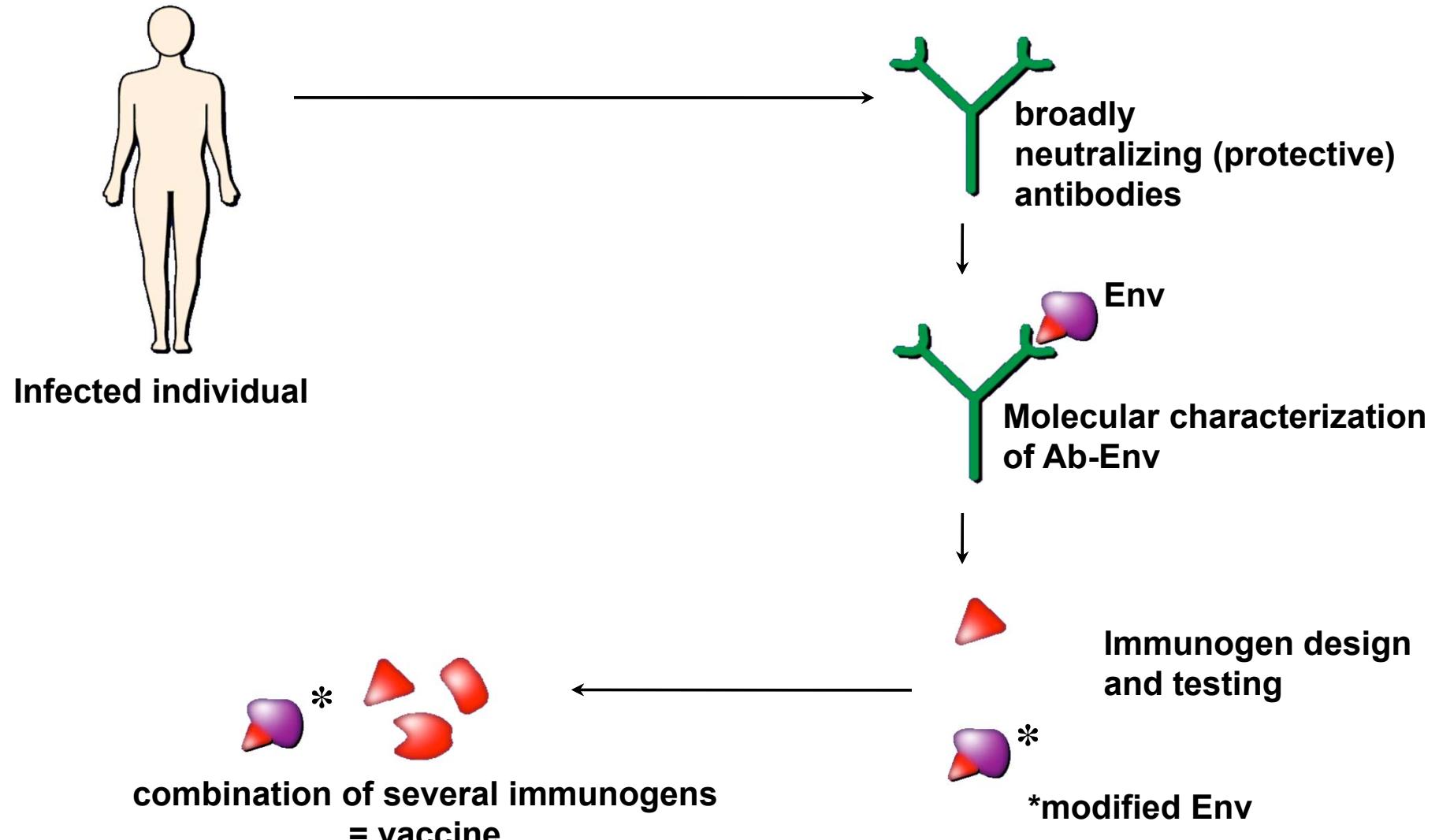
Bill Schief

Professor, The Scripps Research Institute
Director, Vaccine Design, International AIDS Vaccine Initiative
Center for HIV/AIDS Vaccine Immunology and Immunogen Discovery, TSRI
Associate Member, Ragon Institute of MGH, Harvard and MIT

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Vaccine reverse engineering

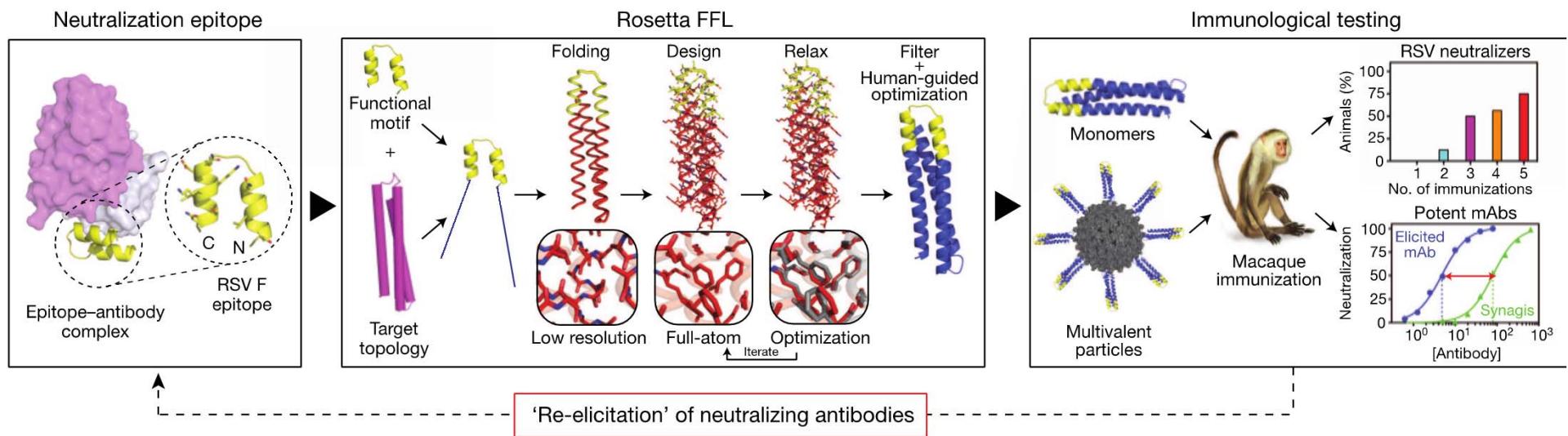


(adapted from Burton, Nat. Rev. Immunol., 2:706, 2002)

Three stories

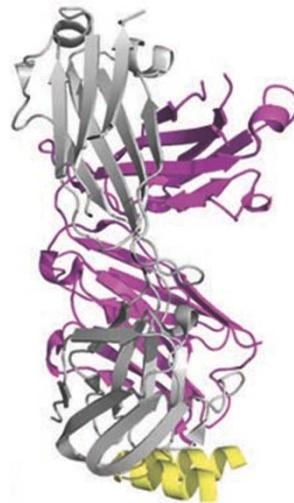
- Proof of principle for epitope-focused vaccine design:
Epitope-scaffolds for motavizumab elicit potent
neutralization of RSV in NHPs
- Toward an HIV vaccine based on the CD4-binding site:
germline targeting to initiate induction of VRC01-class
broadly neutralizing antibodies (bnAbs)
- Proof of principle for elicitation of HIV bnAbs starting from
human germline B cells: vaccine induction of PGT121-class
glycan-dependent bnAbs by germline targeting and
reductionist boosting

Proof of concept for epitope-focused vaccine design: Epitope-scaffolds induce potent neutralization of RSV in NHPs



- A. Potent polyclonal serum neutralizing responses.**
- B. Rhesus mAbs isolated from an immunized macaque are more potent than Synagis**
- C. Rhesus mAbs recapitulate Mota structural specificity.**

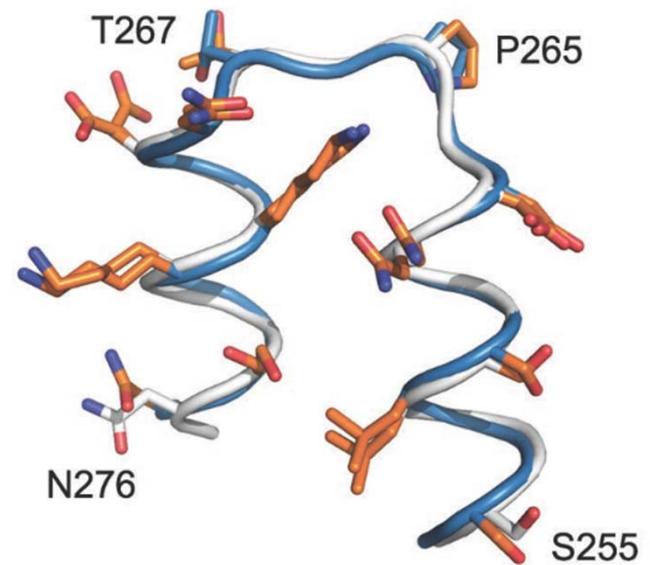
“Re-elicitation” of a neutralizing specificity: Vaccine-elicited mAb targets the same epitope structure as the humanized mAb that guided vaccine design



Guide structure
(Motavizumab+epitope
e)

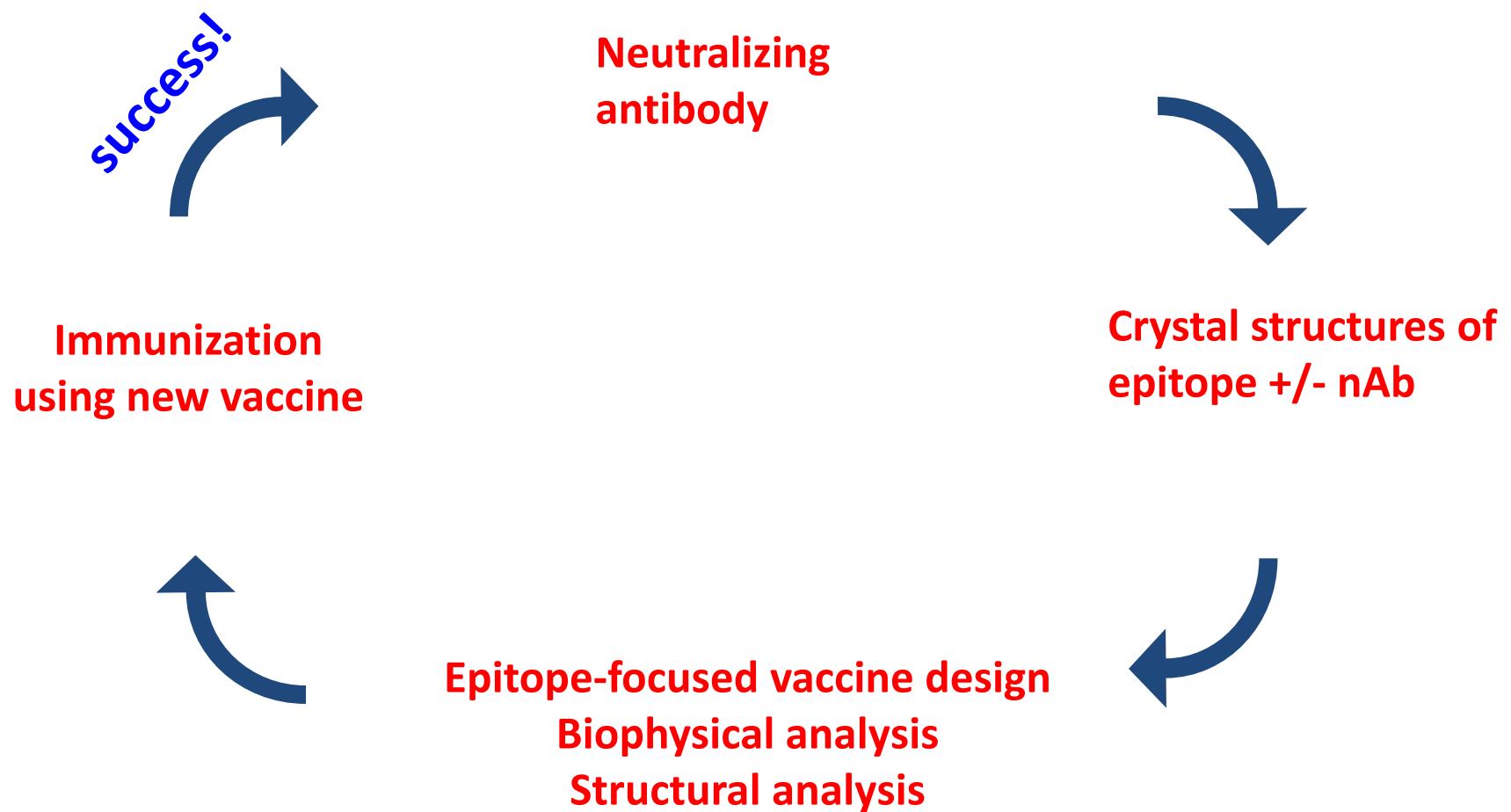


Structure of
vaccine-elicited mAB
(17HD9+epitope)



- ✓ epitope structures superimpose (RMSD = 0.5 Å)
- ✓ 85% of buried side-chains are shared

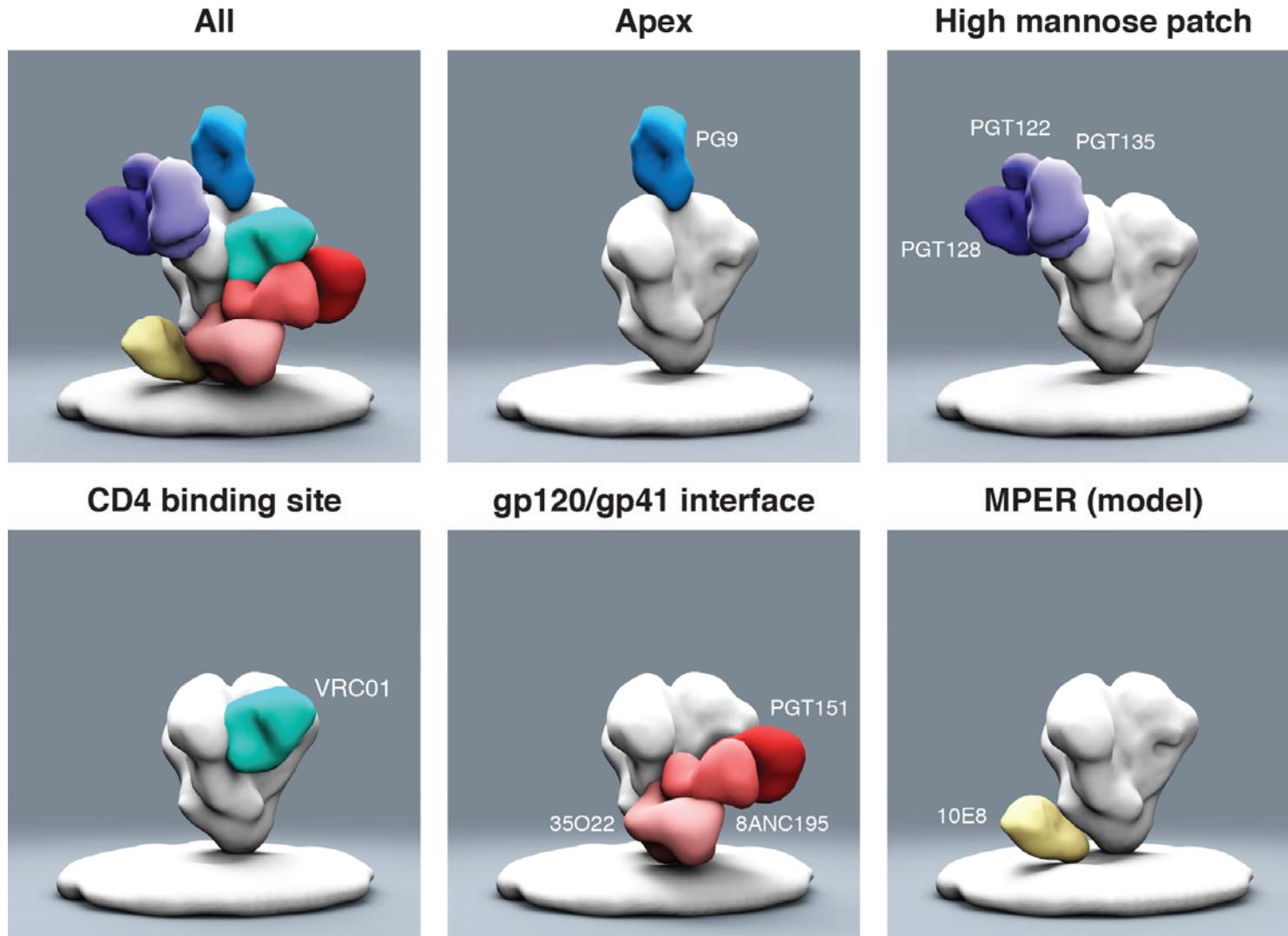
“Closing the loop” of Reverse Vaccine Engineering



Three stories

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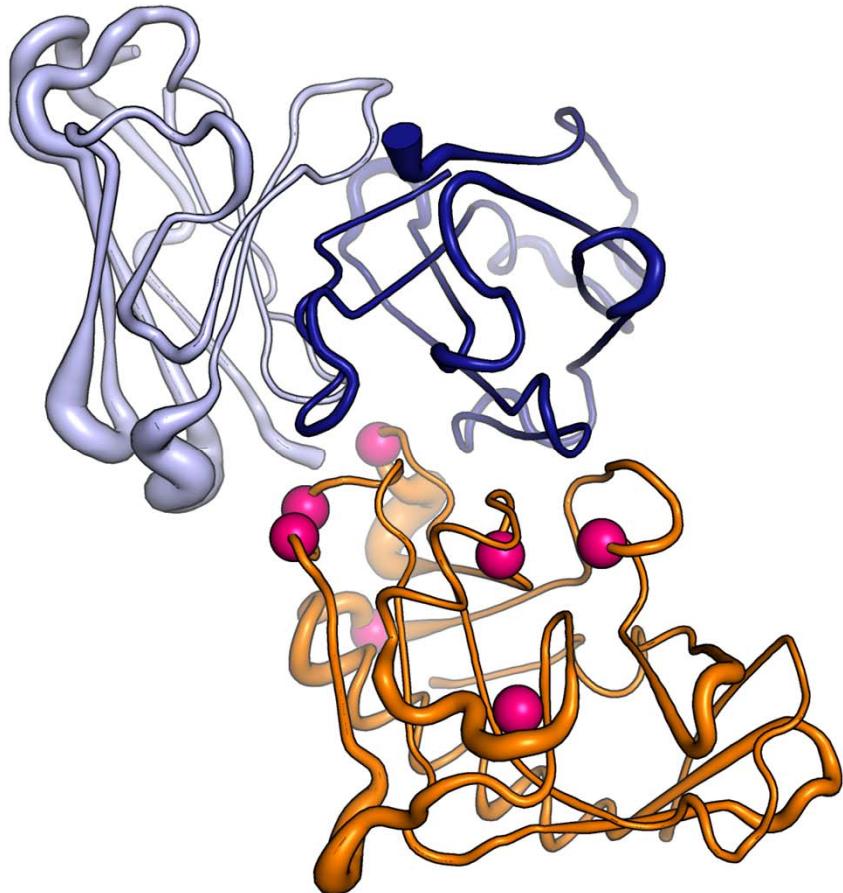
Prototype HIV bnAbs: binding regions



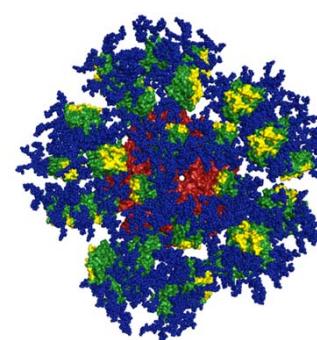
A. Ward & C. Corbaci

Development of VRC01-class Germline-Targeting Immunogen

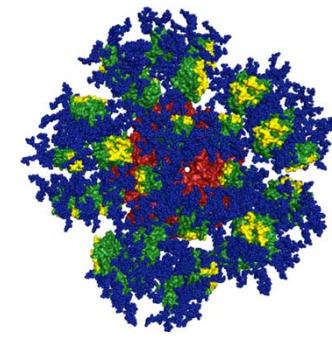
eOD-GT6 bound to GL-VRC01



Self Assembling Nanoparticles

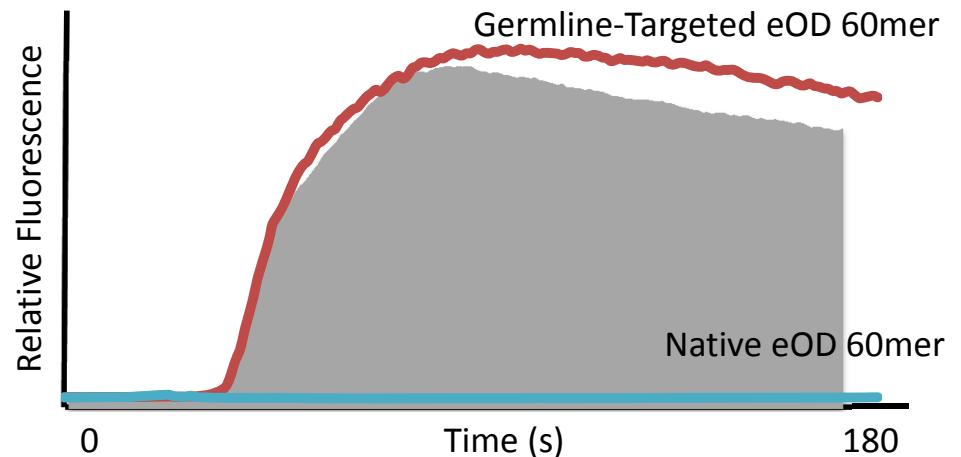


Germline-Targeted
eOD 60mer



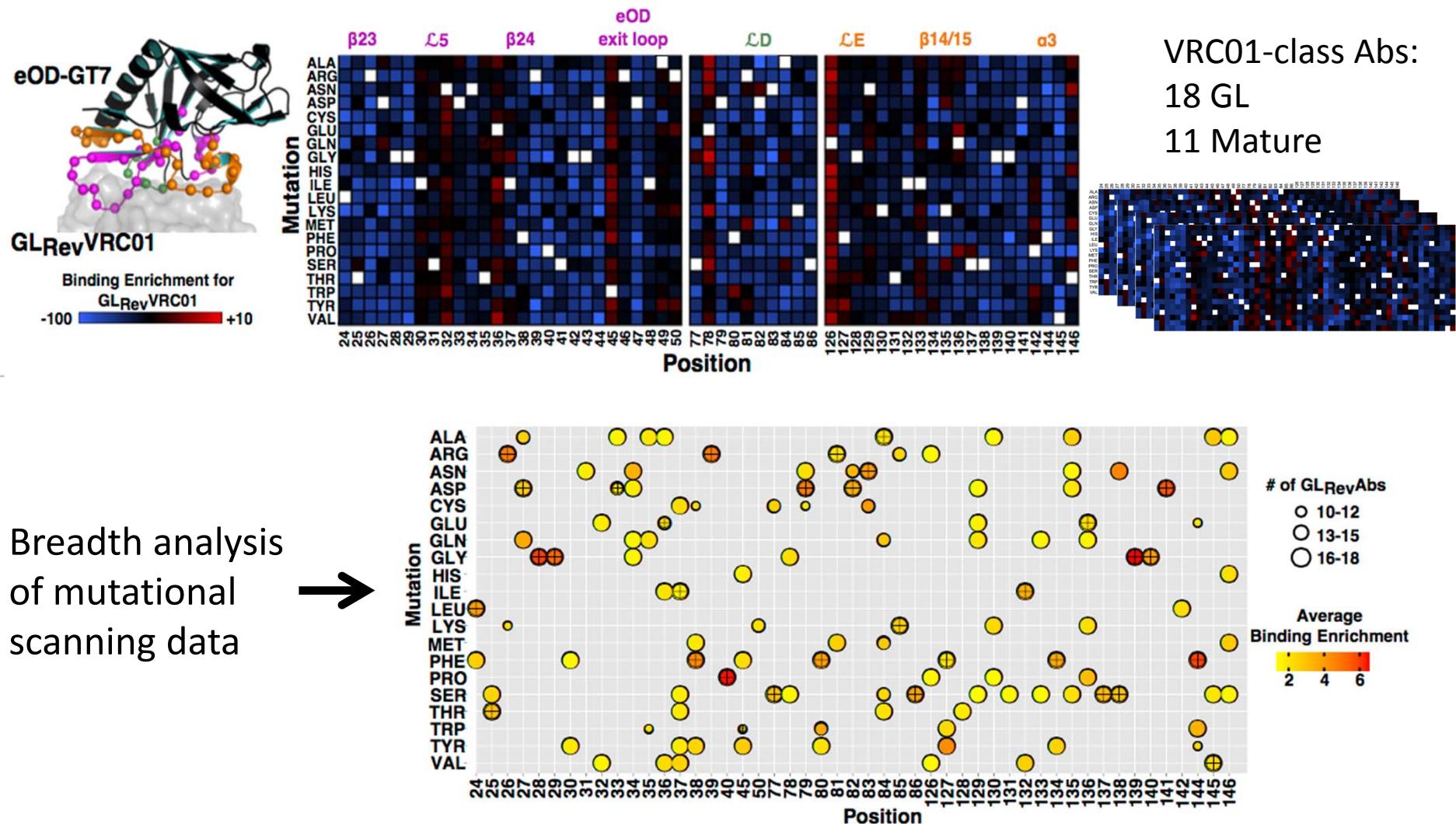
eOD 60mer
(Native CD4bs)

In Vitro Germline VRC01 B Cell Activation

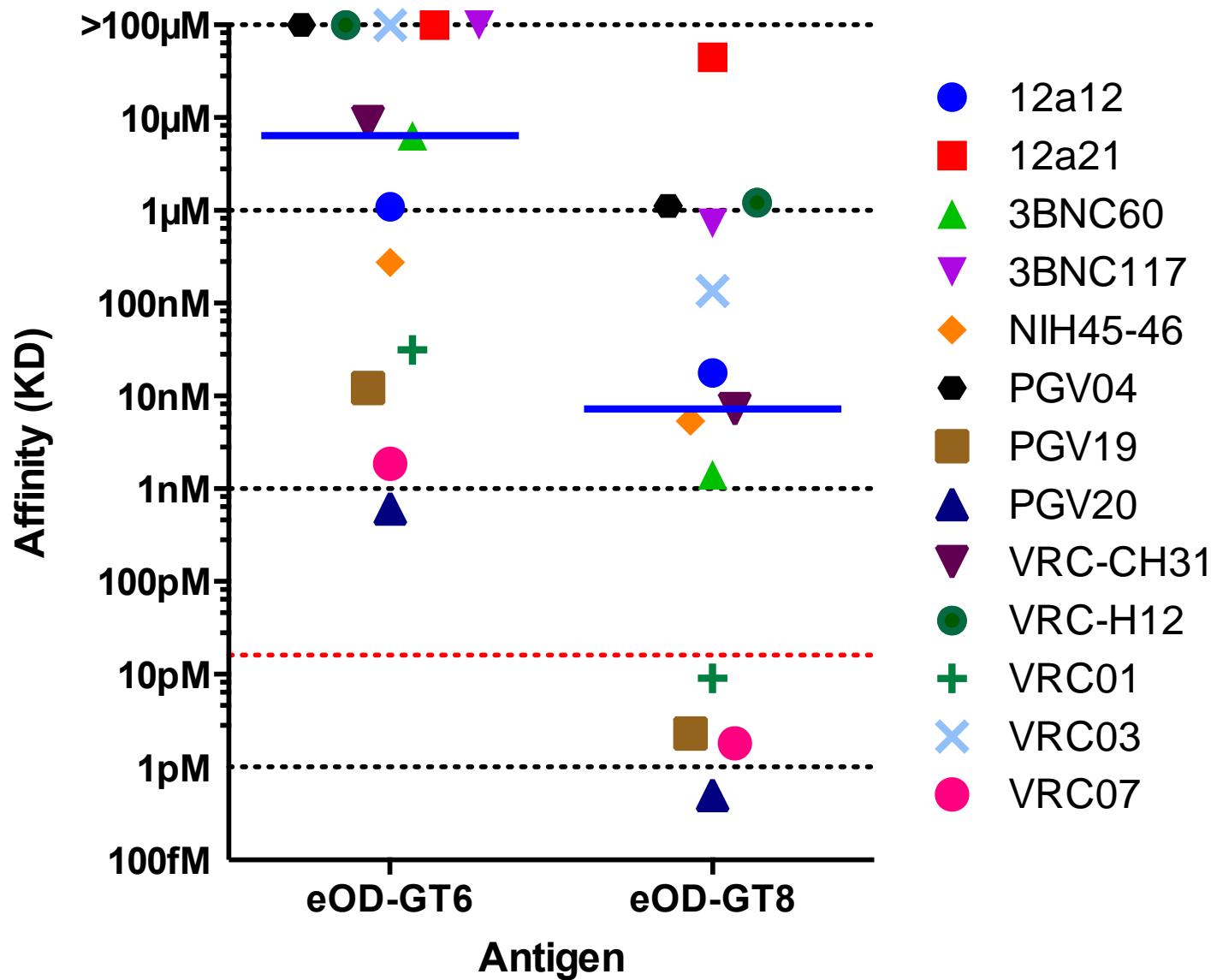


Jardine, Julien, Menis et. al., Science, 2013

Improvement of germline-targeting immunogen by deep mutational scanning and multitarget optimization



eOD-GT8 has improved binding to VRC01-class germline antibodies compared to eOD-GT6



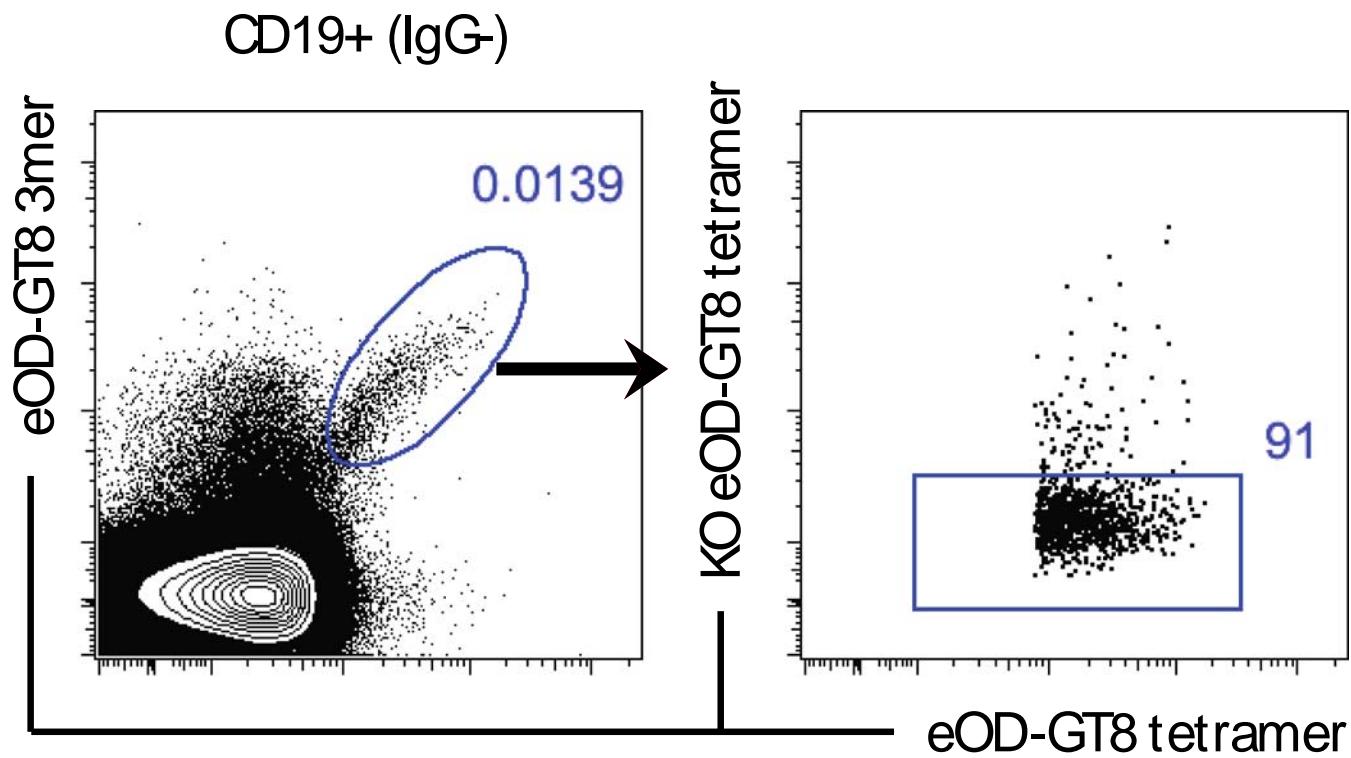
True vs germline-reverted VRC01-class precursors

Design and validation of germline-targeting immunogens has been based on “germline-reverted” precursors that use CDRH3 and CDRL3 loops from mature bnAbs.

This raises several important questions:

- what do “true” VRC01-class human precursors look like?
- How frequently do they occur in HIV-naïve humans?
- Do “true” VRC01-class precursors bind to eOD-GT8?
 - With sufficient affinity to allow B cell activation?

Sorting GT8 specific naïve human B cells



eOD-GT8 isolates VRC01-class precursors from 1 in 2.4 million human naïve B cells

Donor	B cells Screened (millions)	VRC01-class naive B cells	VH1-2 (*02/*03/*04) + 5aa CDRL3
1	1.6	1	
2	2.1	1	
3	0.9	0	
4	5.4	2	
5	0.6	0	
6	0.5	0	
7	1.8	0	
8	14.4	4	
9	7.8	6	
10	4.5	2	
11	7.0	1	
12	5.9	1	
13	1.1	2	
14	6.7	5	
15	1.3	1	
Total	61.6	26	

Frequency: 1 in 2.4 million

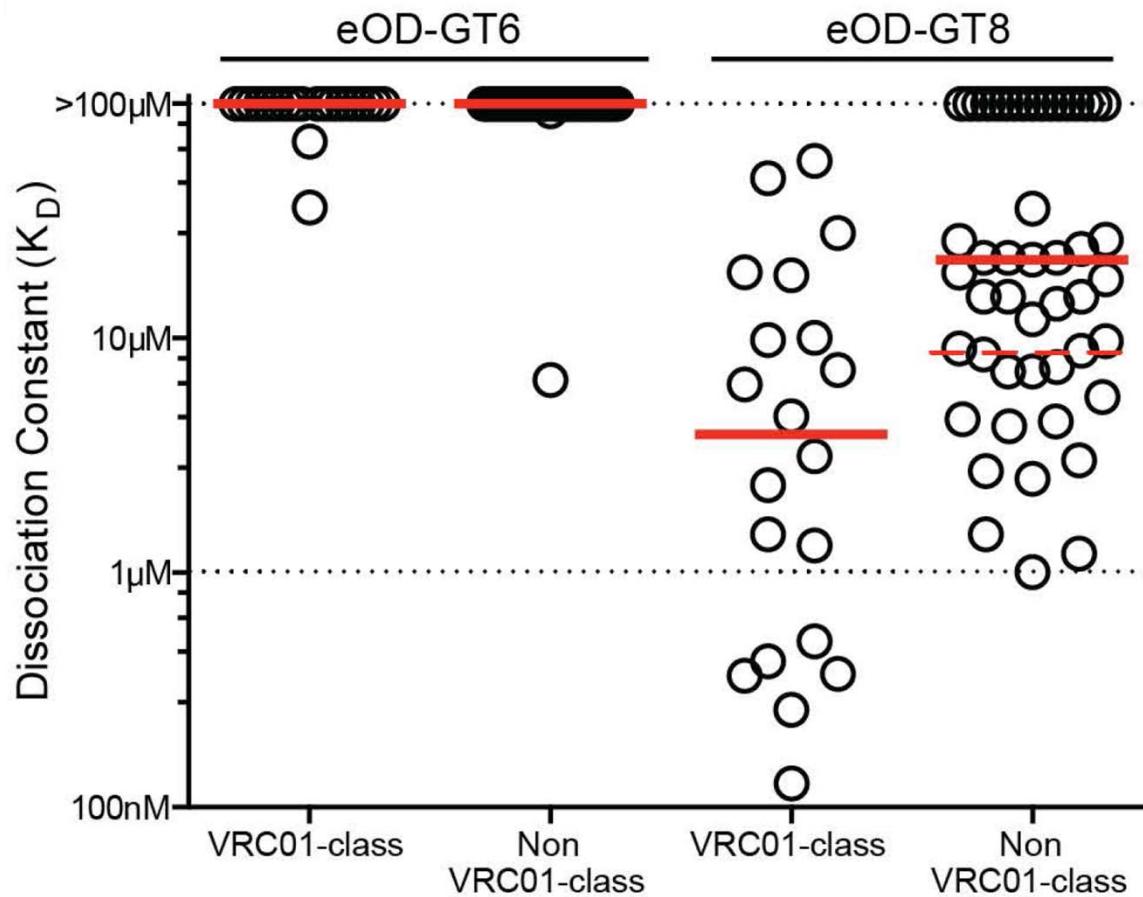
VRC01-class precursors that bind eOD-GT8 are common in humans

- 10^{10} - 10^{11} B cells in adult human
- ~50 million B cells per human lymph node
- 65-75% are naïve B cells
- 96% of humans are hetero/homozygous for VRC01-class VH1-2 alleles (*02, *03, *04)

A frequency of 1 VR01-class precursor in 2.4 million naïve B cells means that:

- nearly all humans have 2700 to 31,000 VRC01-class precursors that bind eOD-GT8
- each human lymph node has ~15 VRC01-class precursors that bind eOD-GT8

eOD-GT8 binds with 0.1-30 μ M affinity to VRC01-class precursors (and has higher affinity for VRC01-class compared to non-VRC01-class)



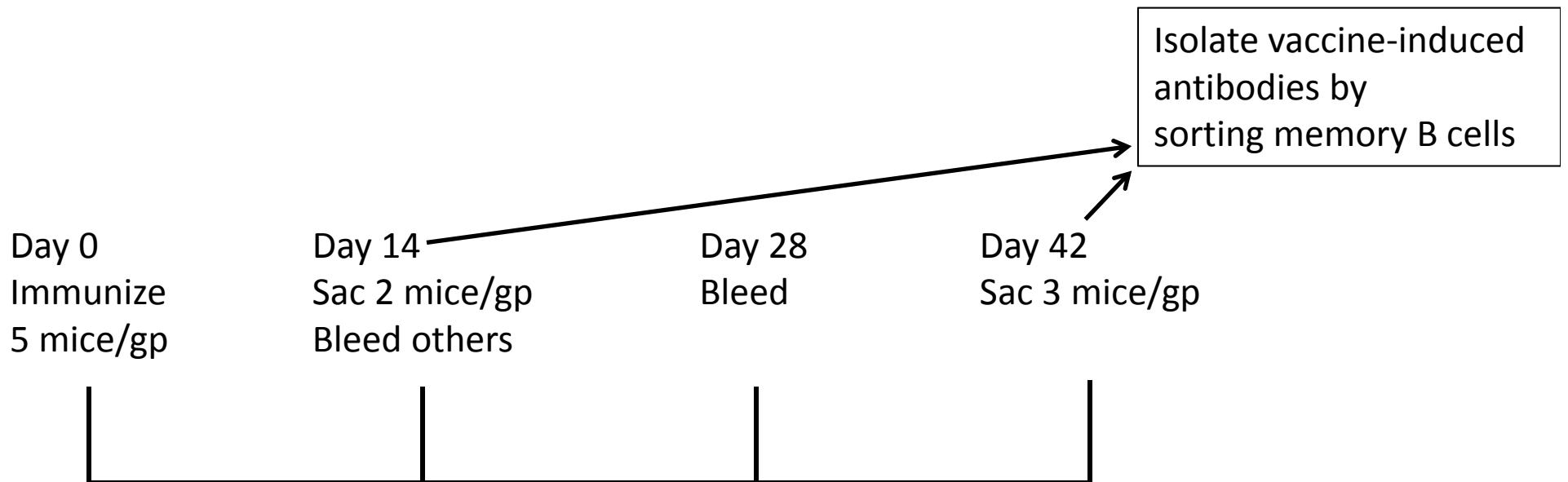
This is promising for *in vivo* activation and VRC01-class memory B cell generation during human vaccination:

- ◆ Batista, Neuberger JEM 1998
- ◆ Dal Porto, Shlomchick JEM 2002
- ◆ Shih, Nussenzweig Nat Imm 2002
- ◆ Paus, Brink JEM 2006

Test of germline-targeting immunogens in VRC01 gH mice

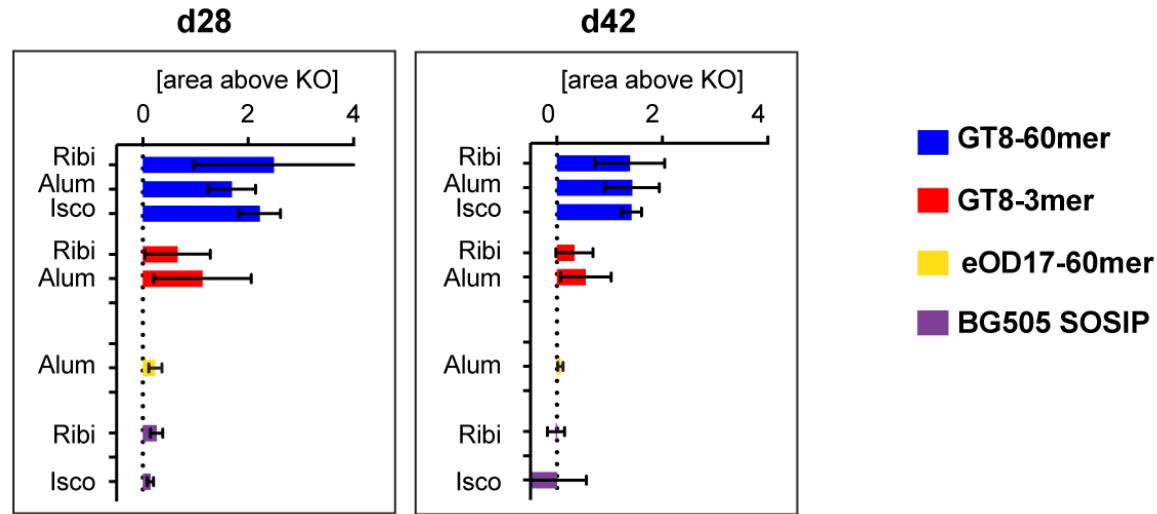
(David Nemazee)

Can eOD-GT8 activate appropriate B cells and select productive mutations, to produce memory B cells that could be boosted by a more native-like immunogen?

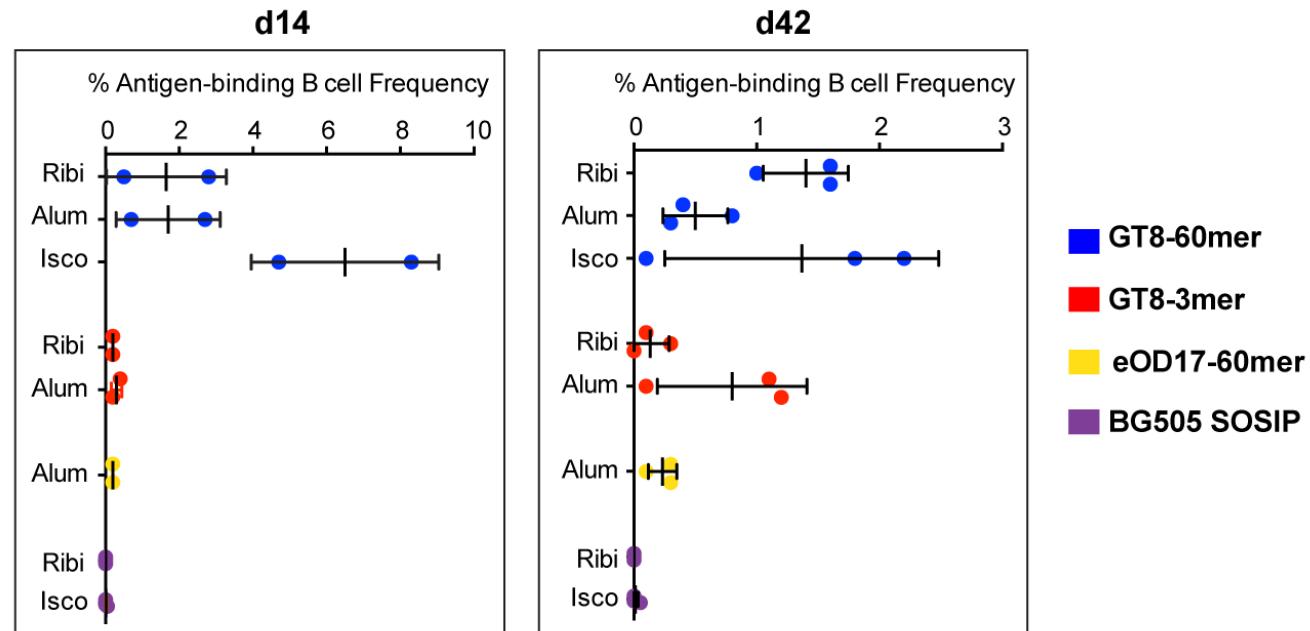


Serum and B cell responses in VRC01 gH mice showed robust responses to the eOD-GT8 60mer

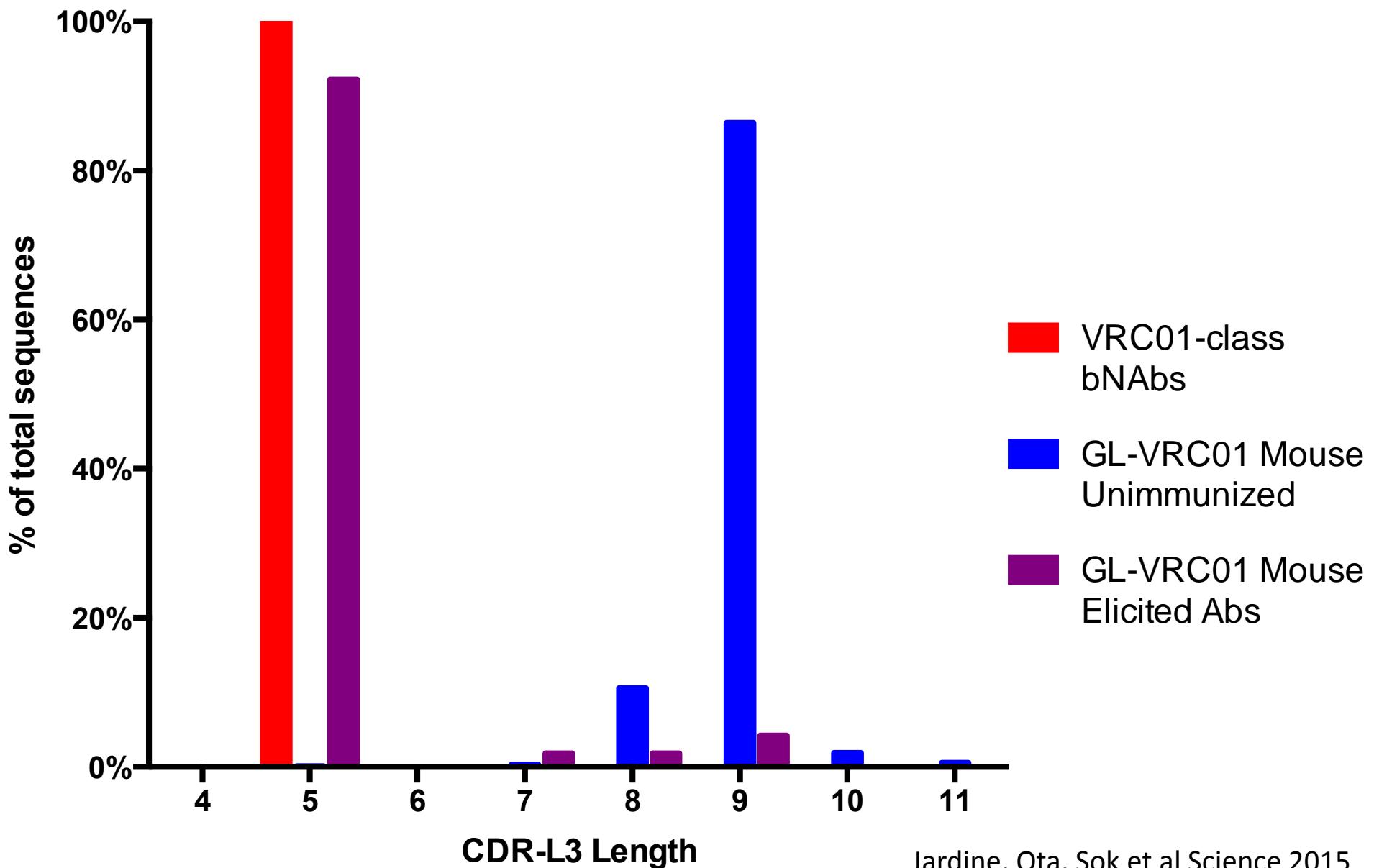
ELISA responses



Epitope-specific B cell frequencies



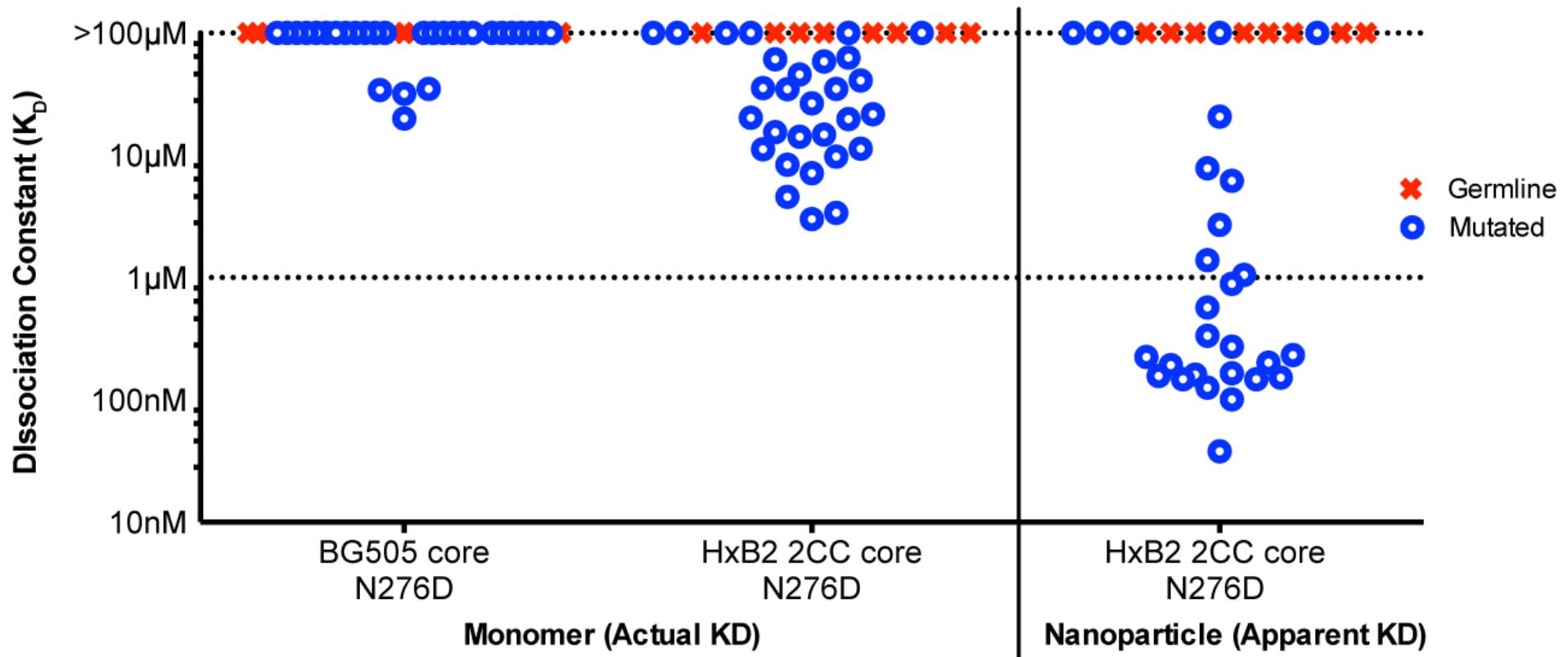
eOD-GT8 60mer reliably elicits Abs with 5AA CDR-L3



Jardine, Ota, Sok et al Science 2015

eOD-GT8 60mer induced Abs bind to near-native CD4bs in candidate boost immunogens

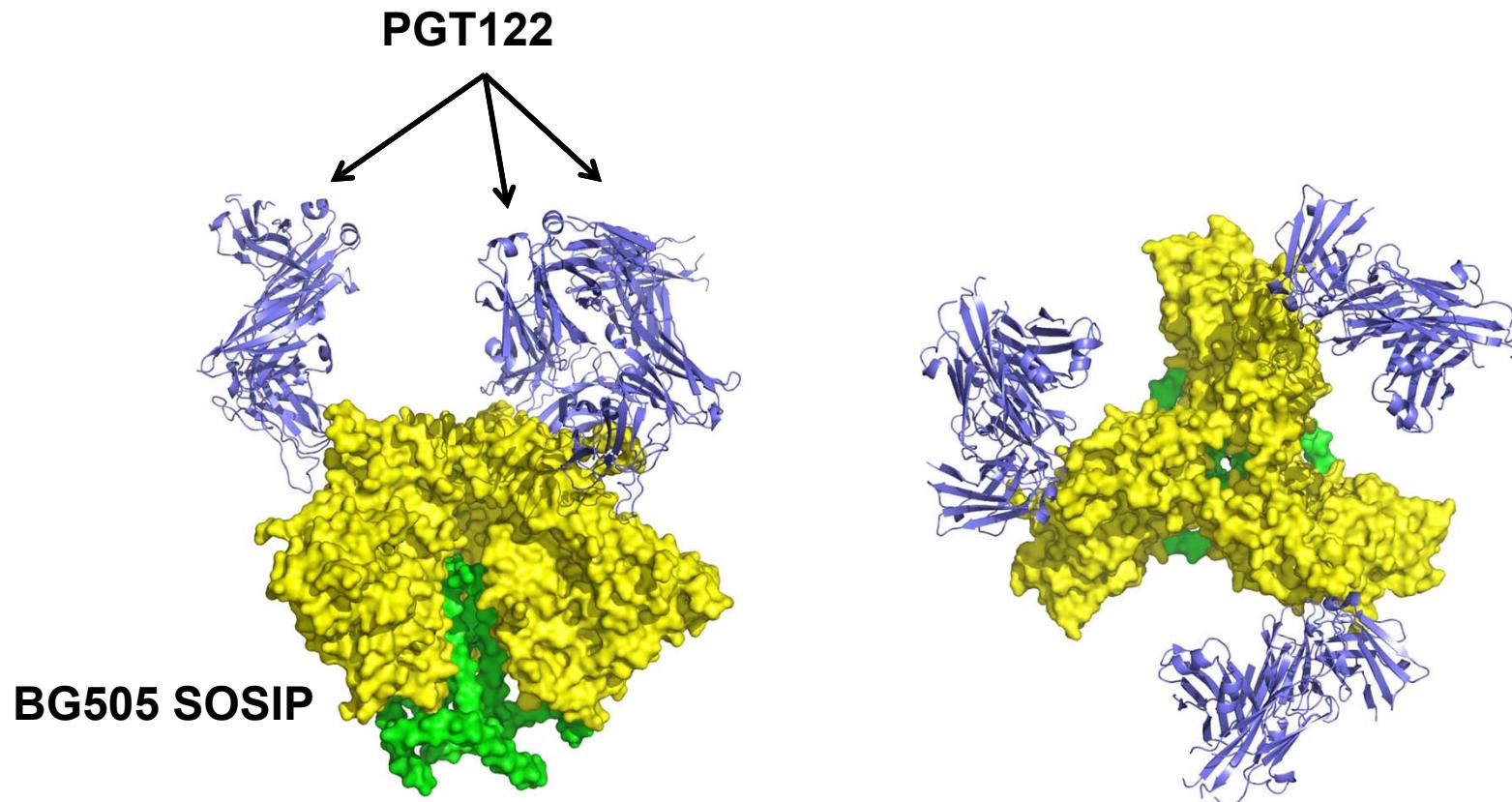
- GT8-induced monoclonal Abs were isolated by sorting of memory B cells from day 42
- Antibody affinities for boost candidates were measured by SPR



Three stories

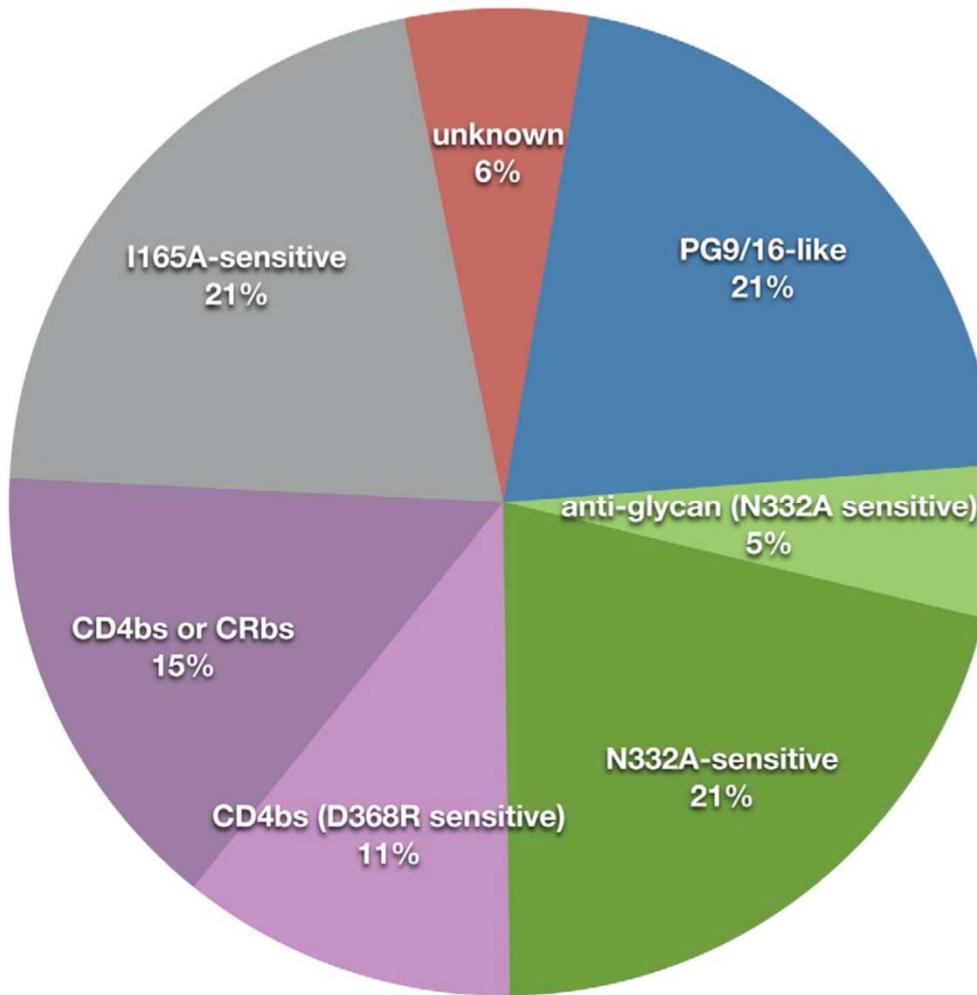
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PGT121-class glycan-dependent bnAbs by germline
targeting and reductionist boosting

PGT121-class interaction with native-like trimer defined



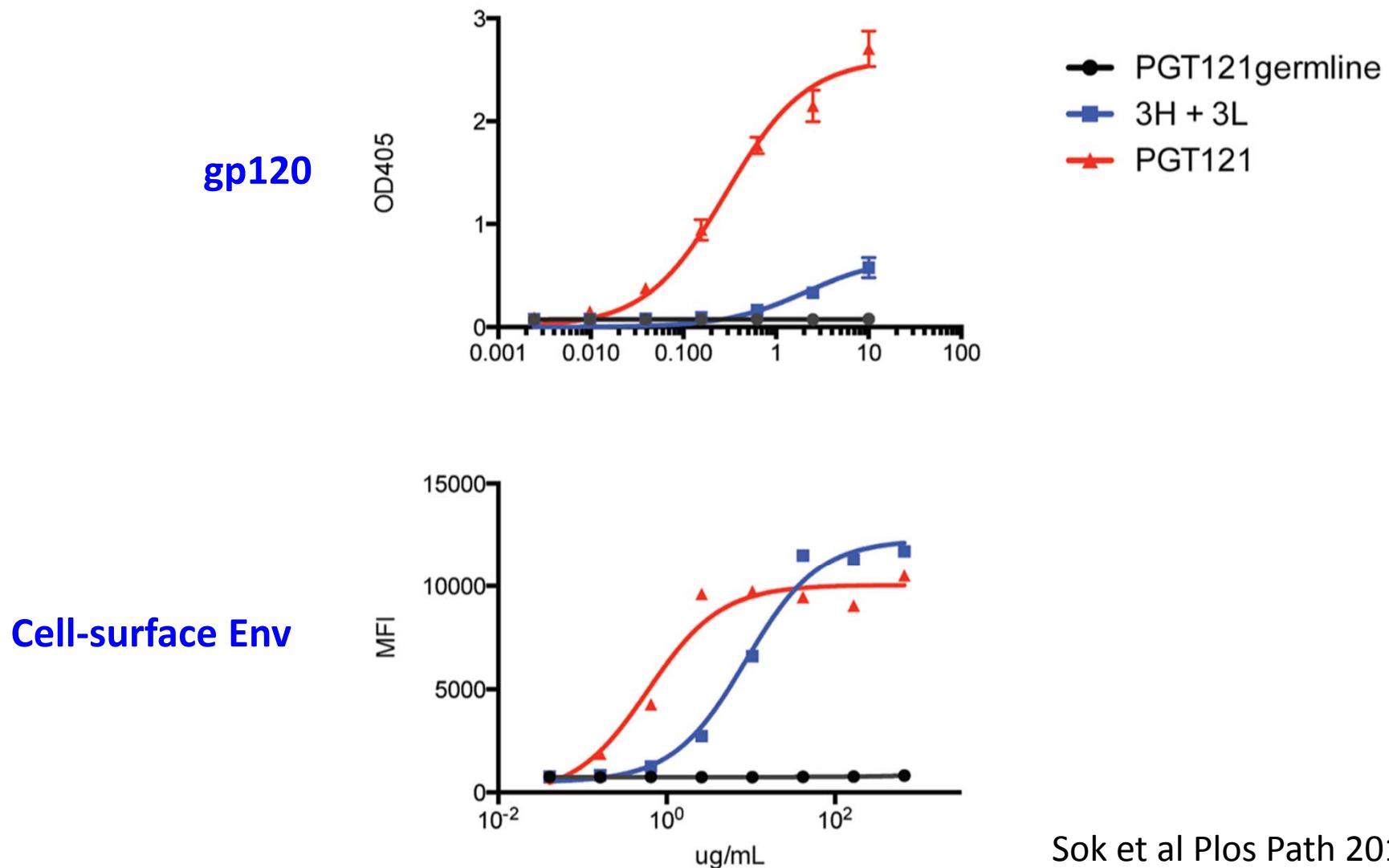
Julien et al Science 2013

PGT121-class bnAbs among the most common from infection



Walker et al Plos Path 2010

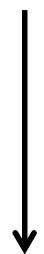
Barrier to elicitation: PGT121 germline-reverted Abs lack detectable affinity for gp120 or cell-surface Env



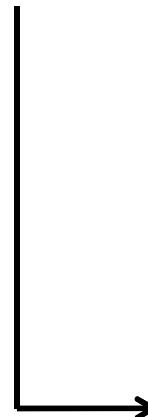
Sok et al Plos Path 2013

Mammalian display/ directed evolution overview

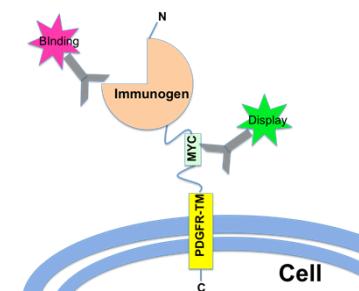
1. Produce lentivirus containing antigen library



2. Transduce 293T cells
Low moi (<0.1)



3. Induce expression (Doxycycline)



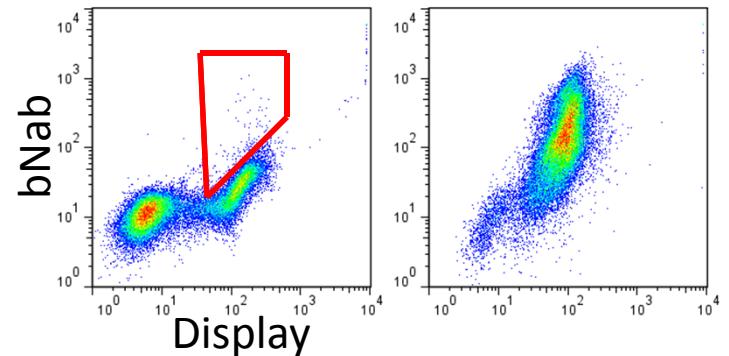
6. Biophysical Characterization



5. Purify genomic DNA, PCR and sequence

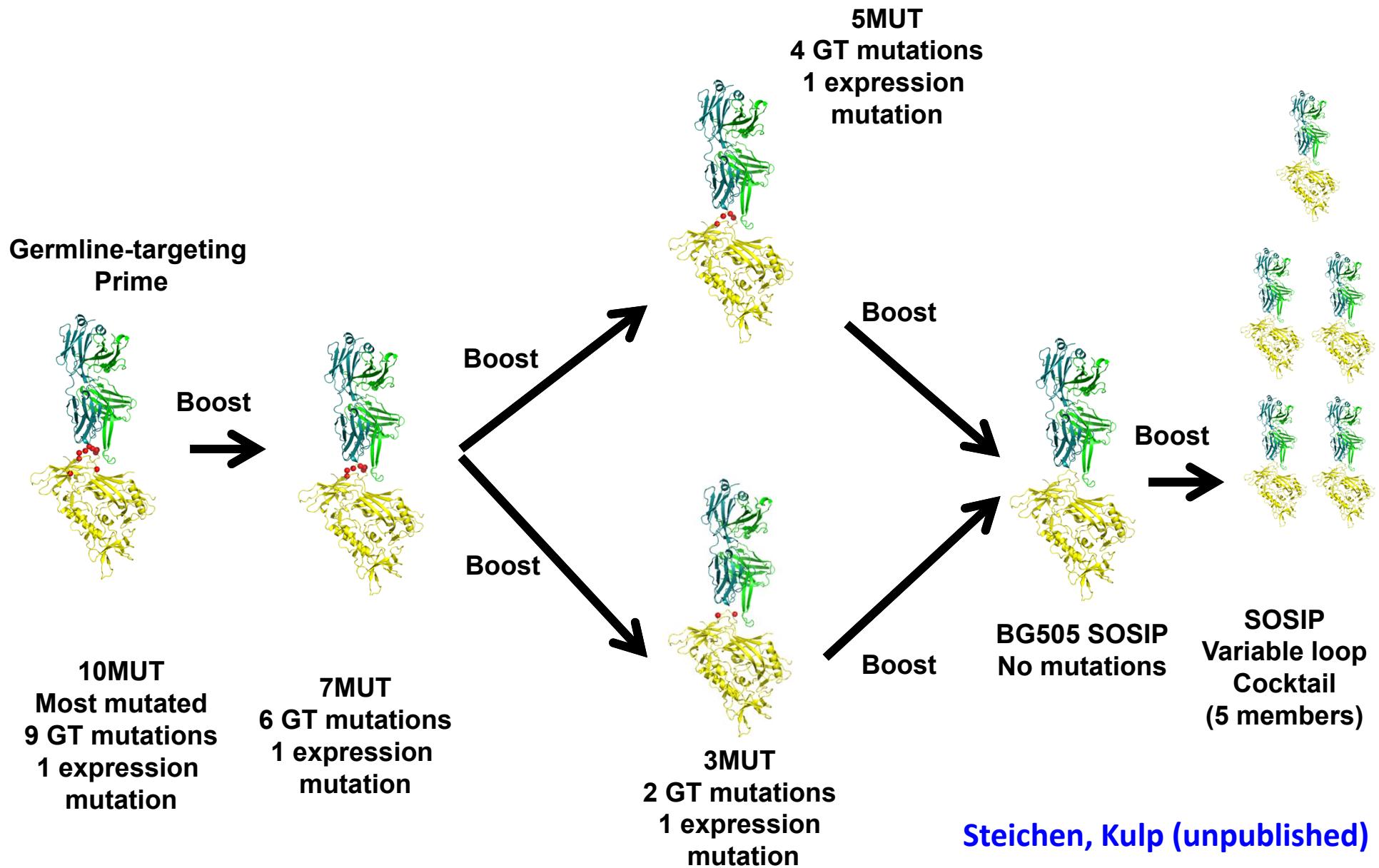


4. Sort cells



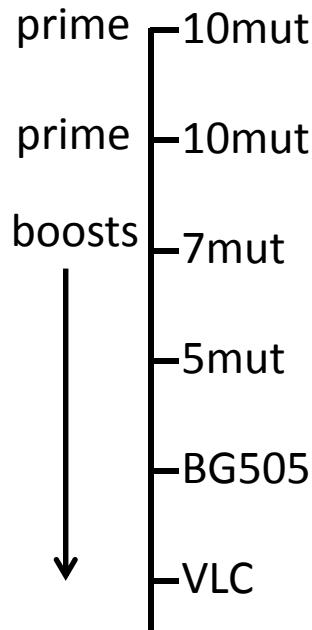
Jon Steichen (unpublished)

Reductionist germline-targeting/boosting strategies to induce PGT121-like bnAbs



Elicitation of tier 2 cross-neutralizing antibodies by reductionist vaccine design in PGT121 gHgL mouse (with Nussenzweig, Burton)

Immunizations
(All SOSIP except 1st)

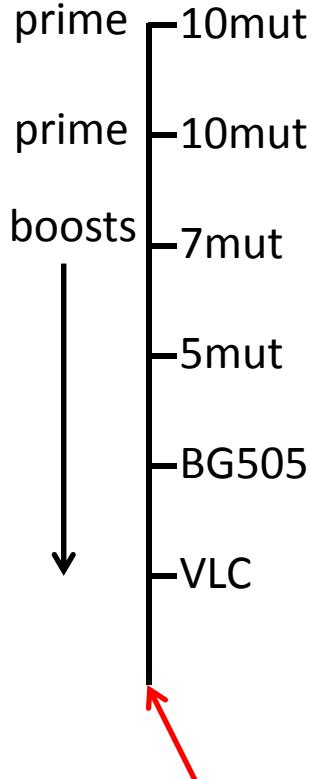


Neut measured
from purified
serum IgG

Escolano, Steichen, et al. (unpublished)

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Specificity of tier 2 cross-neutralization from gHgL mouse tracks closely With PGT121 “ancestor” 3H3L

Virus	clade	MJ6	3H3L	9H3L	PGT121	IC50 ug/mL
6535.3	B	1019	0.043	0.03	0.002	<0.01
TRO.11	B	<25	1.445	0.071	0.005	0.01 - 0.1
CAAN5342.A2	B	1800	0.914	0.236	0.007	0.1 - 1.00
RHPA4259.7	B	<25	>50	0.073	0.015	>1.00
JR-CSF	B	~10	>50	0.277	0.02	
AC10.0.29	B	<25	>50	2.643	0.024	
PVO.4	B	<25	>50	>50	0.098	
SC422661.8	B	<25	>50	>50	0.038	
QH0692.42	B	<25	>50	>50	0.302	
WITO4160.33	B	<25	>50	>50	0.334	
REJO4541.67	B	<25	>50	>50	4.774	
SC05_8C11_2344	B(T/F)	<25	0.386	0.475	0.019	
WEAU_d15_410_50:B(T/F)		<25	1.329	0.3	0.026	
6240_08_TA5_4622	B(T/F)	<25	4.137	0.145	0.033	
1012_11_TC21_3257B(T/F)		<25	>50	0.506	0.003	
1006_11_C3_1601	B(T/F)	<25	>50	14.62	0.002	
1056_10_TA11_182eB(T/F)		<25	>50	2.237	0.004	
1054_07_TC4_1499	B(T/F)	<25	>50	34.74	0.064	
6244_13_B5_4576	B(T/F)	<25	>50	>50	0.061	
62357_14_D3_4589	B(T/F)	<25	>50	>50	2.597	
IAVI C22	C	4429	0.012	0.008	0.002	
DU156.12	C	2877	0.026	0.053	0.004	
HIV-001428-2.42	C	<25	>50	19.74	0.014	
ZM53M.PB12	C	<25	>50	>50	0.001	
Du172.17	C	<25	>50	>50	0.033	
Du422.1	C	<25	>50	>50	0.039	
HIV-16055-2.3	C	<25	>50	>50	0.153	
ZM214M.PL15	C	<25	>50	>50	0.46	
ZM135M.PL10a	C	<25	>50	>50	0.716	
CAP45.2.00.G3	C	<25	>50	>50	1.634	
ZM109F.PB4	C	<25	>50	>50	8.639	
Ce1176_A3	C(T/F)	>5000	0.026	0.078	0.013	
7030102001E5(Rev-)C(T/F)		<25	5.56	1.619	0.009	
1394C9G1(Rev-)	C(T/F)	<25	17.108	1.081	0.264	
Ce704809221_1B3	C(T/F)	<25	>50	11.58	0.025	
246F C1G	C(T/F)	<25	>50	>50	0.041	
ZM247v1(Rev-)	C(T/F)	<25	>50	>50	0.028	
CNE20	BC	2073	0.006	0.005	0.003	
CNE21	BC	<25	2.226	0.256	0.007	
CNE52	BC	<25	>50	3.188	2.045	
CNE19	BC	<25	>50	>50	0.008	
CNE17	BC	<25	>50	>50	7.6	
92RW020	A	1138	0.01	0.012	0.002	
Q23.17	A	>5000	0.007	0.021	0.001	
191084 B7-19	A	<25	38.078	3.994	0.011	
0260.v5.c36	A	<25	>50	15.05	0.53	
0330.v4.c3	A	<25	>50	>50	0.05	
T250-4	CRF02_AG	<25	0.202	0.25	0.001	
235-47	CRF02_AG	<25	>50	>50	0.137	
T251-18	CRF02_AG	<25	>50	>50	29.016	
P1981_C5_3	G	>5000	0.011	0.015	0.001	
X2131_C1_B5	G	<25	8.487	0.224	0.004	
X2088_c9	G	<25	31.256	7.019	0.003	
X1193_c1	G	<25	29.989	11.29	0.016	
X1254_c3	G	<25	1.271	1.832	0.014	
A07412M1.vrc12	D	<25	1.532	0.173	0.009	
6811.v7.c18	CD	3417	0.003	0.001	0.001	
6480.v4.c25	CD	1120	0.033	0.015	0.001	
6952.v1.c20	CD	<25	>50	>50	0.056	
3817.v2.c59	CD	<25	>50	>50	18.888	
3301.v1.c24	C	<25	0.793	0.432	0.008	
3103.v3.c10	ACD	<25	0.062	0.03	0.009	
0815.v3.c3	ACD	<25	0.622	1.549	0.025	

Escolano, Steichen, et al. (unpublished)

Neutralization breadth of best MJ6 mAbs is similar to 3H3L

	MJ6-1	MJ6-2	MJ6-3	3H3L	PGT121	IC50 ug/mL
6535	0.009	0.014	0.026	0.043	0.002	<0.01
92RW020	0.004	0.012	0.022	0.01	0.002	0.01 - 0.1
IAVI C22	0.003	0.008	0.011	0.012	0.002	0.1 - 1.00
Q23	0.005	0.016	0.024	0.007	0.001	>1.00
DU156	0.011	0.024	0.059	0.026	0.004	
P1981	0.004	0.007	0.016	0.011	0.001	
X2088	0.037	0.609	0.316	31.3	0.003	
191084B7	0.712	0.862	>50	38.1	0.011	
JR-CSF	0.334	0.739	>50	>50	0.02	
BG505 T332N	0.55	0.598	>50	0.064	0.026	
T250	>50	>50	>50	0.202	0.001	
HIV-001428	>50	>50	>50	>50	0.014	
PV0.4	>50	>50	>50	>50	0.098	
ZM53	>50	>50	>50	>50	0.001	
CNE19	>50	>50	>50	>50	0.008	
R1166	>50	>50	>50	>50	>50	
MLV	>50	>50	>50	>50	>50	

Conclusions/Outlook

- ◆ A key challenge for HIV vaccine design is immuno-focusing to bnAb epitopes
- ◆ Similar challenges for other antigenically highly variable pathogens such as influenza and hepatitis C viruses, and related challenges for dengue virus
- ◆ RSV scaffold immunogens:
Re-capitulation of Mota neutralization specificity provides proof of principle that epitope-focused vaccine design can achieve immuno-focusing with high precision
- ◆ Vaccines to induce bnAbs against HIV:
Hypothesize that (a) germline-targeting is needed to consistently activate bnAb precursors in vaccine recipients and (b) structure-guided boosting strategies are needed to guide SHM to produce bnAbs.

VRC01 example: Germline-targeting eOD-GT8 60mer has promise to initiate induction of VRC01-class bnAbs. To be tested in humans (IAVI/BMGF). Reductionist boosting strategies to induce VRC01-class bnAbs being tested in various transgenic mice.

PGT121 example: Demonstrated proof of principle for vaccine-induction of HIV bnAbs starting from human germline B cells, by PGT121 germline-targeting and reductionist boosting (using engineered SOSIP trimers) in PGT121 gHgL mice. **A major milestone for HIV vaccine development.** Precursor frequency in humans remains a question.

Acknowledgments– RSV scaffolds

Schief lab

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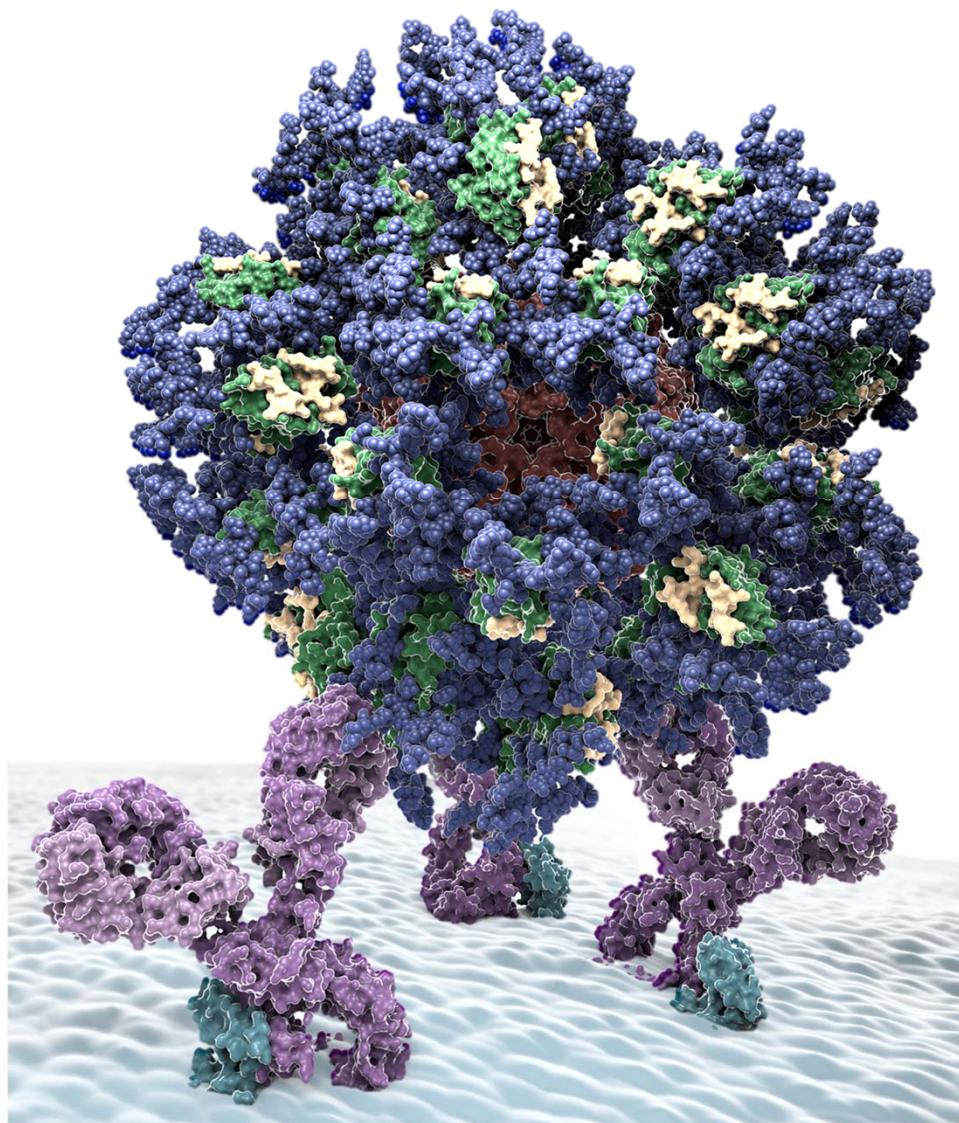
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Burton lab

Laura McCoy

Bryan Briney

Devin Sok

Nemazee lab

Taka Ota

Deepika Bhullar

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